

## Behavioural and physiological effects of electrical stimulation in the nucleus accumbens: a review

K. van Kuyck<sup>1</sup>, L. Gabriëls<sup>2</sup>, P. Cosyns<sup>2</sup>, L. Arckens<sup>3</sup>, V. Sturm<sup>4</sup>, S. Rasmussen<sup>5</sup>, and B. Nuttin<sup>1</sup>

<sup>1</sup> Laboratory of Experimental Neurosurgery and Neuroanatomy, Department of Neuroscience and Psychiatry, Leuven, Belgium

<sup>2</sup> Department of Psychiatry, University Hospital Antwerpen, Edegem, Belgium

<sup>3</sup> Laboratory of Neuroplasticity and Neuroproteomics, Department of Biology, Leuven, Belgium

<sup>4</sup> Department of Stereotaxy and Functional Neurosurgery, Medical University of Cologne, Cologne, Germany

<sup>5</sup> Butler Hospital and Department of Psychiatry and Human Behavior, Brown University School of Medicine, Rhode Island, USA

### Summary

Electrical stimulation (ES) in the brain is becoming a new treatment option in patients with treatment-resistant obsessive-compulsive disorder (OCD). A possible brain target might be the nucleus accumbens (NACC). This review aims to summarise the behavioural and physiological effects of ES in the NACC in humans and in animals and to discuss these findings with regard to neuroanatomical, electrophysiological and behavioural insights. The results clearly demonstrate that ES in the NACC has an effect on reward, activity, fight-or-flight, exploratory behaviour and food intake, with evidence for only moderate physiological effects. Seizures were rarely observed. Finally, the results of ES studies in patients with treatment-resistant OCD and in animal models for OCD are promising.

**Keywords:** Neuromodulation; electrical stimulation; behaviour; nucleus accumbens; ventral striatum; review.

### Abbreviations

5-HT serotonin; 6-OH-DA 6-hydroxydopamine; AMY amygdala; DA dopamine; DAergic dopaminergic; DOPAC 3,4-dihydroxyphenylacetic acid; DTγE (Des-Tyr<sup>1</sup>)-γ-endorphin; ES electrical stimulation; FCV fast cyclic voltammetry; GABA gamma-aminobutyric acid; HC hippocampus; HVA homovanillic acid; ICSS intracranial self-stimulation; LH lateral hypothalamus; MD mediodorsal thalamic nucleus; MDMA methylenedioxymethamphetamine; MFB medial forebrain bundle; NA noradrenaline; NACC nucleus accumbens; SN substantia nigra; SNc substantia nigra pars compacta; SNr substantia nigra pars reticulata; VP ventral pallidum; VTA ventral tegmental area.

### Introduction

Some patients with obsessive-compulsive disorder (OCD) are treatment-refractory to conventional behavioural therapy and/or pharmacological treatment. Part of these patients may benefit from a neurosurgical lesion

in a specific brain target [36, 46]. In one of those brain targets, the anterior limbs of the internal capsule [60, 67], we demonstrated that high frequency ES was also therapeutically effective [77]. In contrast to neurosurgical lesions, electrical brain stimulation is a reversible technique, which is a major advantage in case severe side effects occur. Moreover, in Parkinson's disease ES has a lower rate of side effects compared to lesioning with thermocoagulation [103].

Although the clinical outcome of ES in the anterior limbs of the internal capsule is satisfactory, high voltage levels are necessary. Hence, the battery lifetime is limited to 4–12 months requiring frequent exchange of the batteries under local anaesthetic, limiting the comfort of the patient. One of the strategies to surpass the high energy consumption is to search for other brain targets that yield the same or even better therapeutic results with a lower voltage.

A possible new target for ES in patients with treatment-refractory OCD might be the nucleus accumbens (NACC) [1, 117], which participates as the antero-ventral part of the ventral striatum in the cortico-striato-pallido-thalamo-cortical circuitry. Functional brain imaging studies indicate that this circuitry is involved in OCD [100]. Additional evidence for the involvement of the NACC in OCD comes from stereotactic lesioning studies in the anterior limbs of the internal capsule. Lesioning of the ventro-caudal part of the internal capsule was imperative for successful treatment. It is likely that such a lesion also affects the NACC [67, 110]. The

current article reviews the reported behavioural and physiological effects of ES in the NACC of humans and different mammalian species.

## Methods and results

We performed a computer-aided search of Pubmed using the keywords 'nucleus accumbens', 'ventral striatum' and 'ES' and selected articles dealing with the behavioural and physiological effects of ES in the NACC. In addition, we searched the reference lists of these selected relevant articles. Because we cannot read or comprehend Chinese lan-

guage one article was rejected [53]. The results of our search are described in the following sections.

## Anatomy of the NACC

### Core and shell NACC

The NACC has been subdivided in a core and shell subregion based on cytoarchitectonic and neurotransmitter characteristics and differences in afferent and efferent connections [142]. The shell is situated in the medial

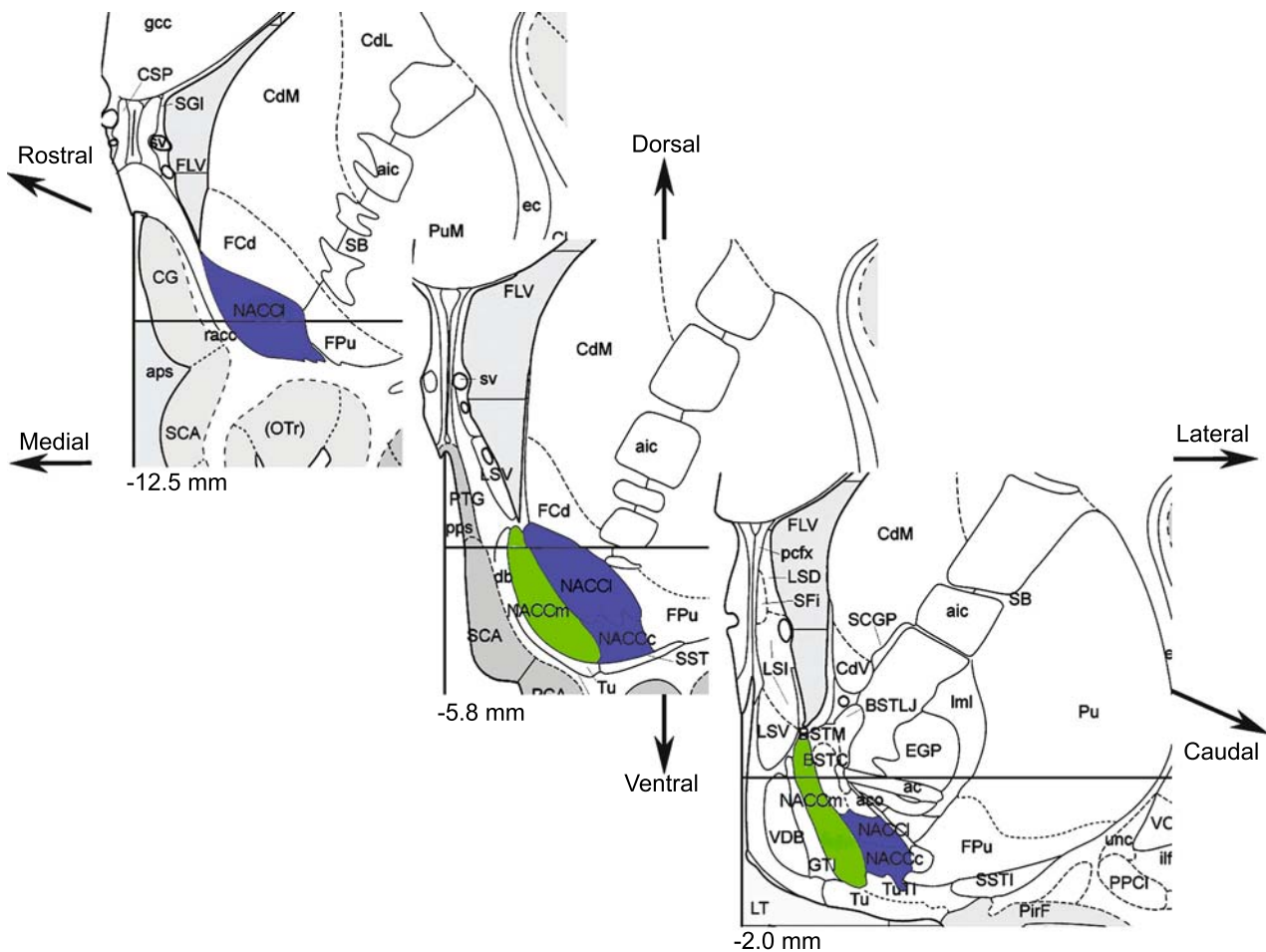


Fig. 1. NACC core and shell: Graphical representation of coronal sections through the human NACC core (blue) and shell (green) and its surrounding structures (modified from the atlas of Mai *et al.* 63). The distance from the midpoint of the anterior commissure at the midline is denoted below each coronal section. *ac* Anterior commissure; *aco* anterior commissure, olfactory limb; *aic* anterior limb of internal capsule; *aps* anterior parolfactory nucleus; *BSTC* bed nucleus of the stria terminalis, central division; *BSTLJ* bed nucleus of the stria terminalis, lateral division, juxtacapsular part; *BSTM* bed nucleus of the stria terminalis, medial division; *CdL* lateral caudate nucleus; *CdM* medial caudate nucleus; *CdV* ventral caudate nucleus; *CG* cingulate gyrus; *CSP* cavity of septum pellucidum; *db* diagonal band; *ec* external capsule; *EGP* external globus pallidus; *FGd* caudate fundus region; *FLV* frontal horn of the lateral ventricle; *FPu* putamen fundus region; *gcc* genu of the corpus callosum; *GTI* great terminal island; *lml* external medullary lamina of the globus pallidus; *LSD* dorsolateral septal nucleus; *LSI* intermediolateral septal nucleus; *LSV* ventrolateral septal nucleus; *NACCc* accumbens nucleus, central (subventricular) part (core); *NACCm* accumbens nucleus, medial (subventricular) part (shell); *OTr* olfactory trigone; *pcfx* precommissural fornix; *PirF* (pre-)piriform cortex, frontal area; *PPCI* (pre-) piriform claustrum; *pps* posterior parolfactory sulcus; *PTG* paraterminal gyrus; *PuM* medial putamen; *racc* radiation of corpus callosum; *SB* striatal cell bridges; *SCA* subcallosal area; *SCGP* supracapsular part of the globus pallidus; *SFi* septofimbrial nucleus; *SGI* substantia nigra; *SSTI* subthalamic terminal island; *sv* septal vein; *Tu* olfactory tubercle; *TuTI* tubercular terminal island(s); *unc* uncinatus fasciculus; *VDB* vertical limb of the diagonal band

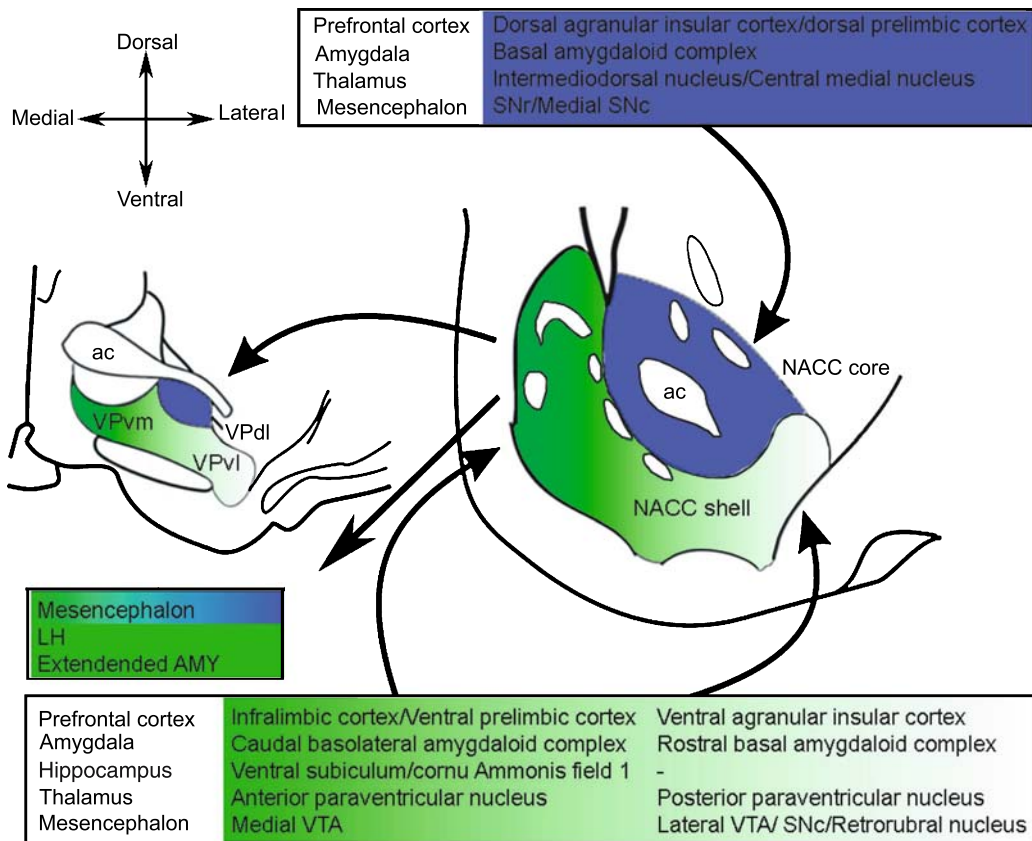


Fig. 2. Neuroanatomical connections of the NACC: The topographical organisation of the main afferents and the VP efferents of the NACC core and shell is represented in a transverse section through the NACC (right) and the VP (left) (modified from Ref 15). *ac* Anterior commissure; *NACC* nucleus accumbens; *VPdl* dorsolateral ventral pallidum; *VPvl* ventrolateral ventral pallidum; *VPvm* ventromedial ventral pallidum

and ventral part of the caudal two thirds of the NACC and encompasses the core, situated laterally in the NACC [19] (Fig. 1). The NACC core is connected to the extrapyramidal motor system and the NACC shell to limbic brain areas mediating emotional processes. In addition to the NACC shell and core, some authors have recognized a rostral pole [152].

#### Main afferent and efferent projections

We shortly summarize the main connections of the NACC (see Fig. 2) but refer to other reviews for in depth expositions of the NACC neuroanatomy in the rat [37] and nonhuman primates [39]. For a topographical organisation of the connections of the NACC core and the shell, we refer to the Fig. 2 (obtained with permission from 37).

The NACC receives mainly glutamatergic projections from the amygdala (AMY) [92], hippocampus (HC) [17, 61, 121, 135, 150], thalamus [6, 9] and prefrontal cortex (PFC) [11, 13, 23, 34, 71, 72, 105, 113] and a dopaminergic (DAergic) projection from the mesence-

phalon, i.e. ventral tegmental area (VTA) and substantia nigra (SN) [22, 85]. The major efferent projection from the NACC terminates in the ventral pallidum (VP) and is principally gamma-aminobutyric acid (GABA)ergic [14, 41, 137, 143]. The ventral pallidum, in turn, projects strongly to the substantia nigra pars compacta (medio-lateral part) as well as to the limbic part of the subthalamic nucleus and its extensions into the local hypothalamus [38]. In addition, the NACC provides a recurrent projection to the VTA and SN in the mesencephalon [41]. An important difference between NACC core and shell is the efferent projection from the NACC shell to the lateral hypothalamus (LH) and the extended AMY, which does not exist in the NACC core [41].

#### Signal processing in the NACC

##### 'Up' and 'down' membrane potential states

More than 90% of the projection neurons in the NACC are medium-sized neurons with spines on their dendrites [4, 12, 79]. These neurons show membrane

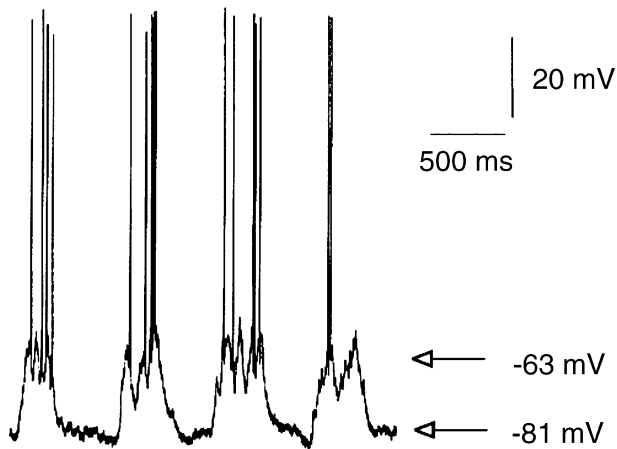


Fig. 3. Typical firing pattern of NACC neurons: Typical firing pattern of most NACC neurons exhibit 'up' and 'down' states in their membrane potential. Action potential firing is only observed during these 'up' events (reprinted from O'Donnell, 1999 [46])

potential shifts ( $\pm 20$  mV) from a negative 'down' state to a depolarized 'up' state [141] (Fig. 3), which are regulated by inputs from the HC [80] and the VTA [35]. The potential shifts bring the membrane potential during 100–1000 ms close to the firing threshold enabling other inputs (e.g. from the PFC) to evoke action potentials in the NACC (see Fig. 3). Hence, afferents from the HC and the VTA are able to gate other inputs. In accordance, slow frequency firing was recorded in the NACC during the 'up' state membrane transitions, taking place at a frequency less than 1 Hz [35].

#### Neuronal ensembles

The transitions in membrane potentials to the 'up' state occur synchronously in ensembles of NACC neurons, rather than in the global NACC or in single NACC neurons [81]. Many of the afferent projections from the abovementioned brain areas converge their input on single NACC neurons and their dendrites within these ensembles [18, 106]. Based on the topographical organisation of afferent connections to the NACC, each of these ensembles integrates different inputs. Likewise, the NACC ensembles relay the input to distinct output areas upon activation.

#### Presynaptic modulation of input

In addition to directly modulating the neuronal activity of NACC neurons, afferent projections also modulate the input of other afferents. For instance, VTA activation of D2 receptors on terminals of hippocampal afferents to the NACC, enhances the excitability of these neurons [136].

### Behavioural effects of ES in the NACC

ES in the NACC has an effect on a wide range of behaviours, which will be discussed in the following sections. ES in the NACC has rewarding properties: animals with an electrode in the NACC will perform *self-stimulation (intracranial self-stimulation)* to apply electrical pulses in the NACC. In addition, ES in the NACC influences activity, fight-or-flight behaviour, exploratory behaviour and food intake. The effect of ES in the NACC on OCD symptoms in animal models and humans will be discussed in the following sections. Finally, the risk for seizures will be evaluated.

#### Reward – intracranial self stimulation (ICSS)

In 1954, Olds and Milner discovered that rats perform an operant task to apply trains of electrical pulses in the septal area and other regions of the brain [82]. This behaviour, called intracranial self-stimulation, has rewarding properties probably by activation of neural systems, which mediate natural rewards like food intake [126]. Therefore, ICSS has been used in models for depression to quantify the ability to experience pleasure. In these models, a decrease in ICSS rate or an increased threshold for ICSS is indicative for depression. The clinical effect of supposed new antidepressants was predicted in the animal models by evaluating the effect on ICSS rate or threshold. The NACC is one of the brain targets where it is possible to induce ICSS (for references, see below). In the NACC, ICSS is accompanied by a highly stereotypic backing away from the lever, sniffing, licking and digging [69].

#### Regional differences in ICSS rate within the NACC

The rate of ICSS in the NACC may be more than 20 per minute [95]. It depends on the NACC subregion where the electrode tip is located. In mice, differences in ICSS rates were noticed with rostro-caudal and dorso-ventral gradients. In the dorsal NACC, ICSS rates were very high rostrally but decreased more caudally. To the contrary, in the ventral NACC, ICSS rates were almost absent rostrally whereas good ICSS rates were observed caudally, with even higher values than in the dorsal NACC at this level. In regions intermediate between dorsal and ventral NACC, there was generally a good responding with higher ICSS rates caudally than rostrally [147]. In another study, a medio-lateral gradient was observed with higher ICSS rates in the medial NACC [89]. Only in one study,



in Rhesus monkeys, no ICSS was observed in the NACC [133].

#### *ICSS in the NACC versus ICSS in other brain targets*

There are also differences in ICSS rates between the NACC and other brain regions. ICSS rates in the NACC are lower than in the LH [123, 93], the medial forebrain bundle (MFB) [99, 144], the VTA [69, 123], the PFC [109] and the substantia nigra (SN) [8, 90, 144] but higher than in the caudatoputamen, the AMY and the olfactory tubercle [89]. One study found no difference in ICSS rate in the NACC versus the SN [21]. Except for the above-mentioned studies, no other comparisons in ICSS rate between NACC and other brain targets were reported. These differences in ICSS rates between brain areas, however, may depend on the duration of the test period. For instance, it was reported that rats self-stimulate at the same rate in the NACC like in the LH but that rats with an ICSS electrode in the NACC need more days to achieve these equal ICSS rates [47]. Differences in the number of days to acquire ICSS were also observed within the NACC itself. ICSS was faster acquired in dorsomedial anterior versus posterior NACC subregions [147].

#### *Stimulation parameters*

In the ICSS paradigm, animals work to obtain *trains of electrical pulses* (train duration: 0.1–1 s [99, 149] in a rewarding brain area. In the NACC, animals will respond with higher ICSS lever pressing rates if the *frequency* of these pulses in the train is increased, with a plateau at approximately 60 Hz [123]. Also changes in pulse parameters influence the ICSS rate. The ICSS rate increases as a function of *current intensity* [8, 47, 69, 109] until a plateau is reached [90]. In rats with a lesion in the VTA (and consequently with a destruction of DAergic input to the NACC), ICSS decreased at higher current intensities [21]. As far as we know, there are no studies in which the effects of different *pulse widths* were compared. In most of the studies, pulse widths between 0.2 and 0.3 ms were used [59, 148]. Typical pulse *waveforms* used to induce ICSS in the NACC were monophasic [90] and biphasic square [148] wave pulses, as well as sine wave pulses [89].

#### **Influence of stress on ICSS in the NACC**

The NACC is not only involved in processing reward but also in mediating stress responses [130] (see also

below: Fight-or-flight). Exposure to acute stress results in an enhanced DA and serotonin (5-HT) release in the NACC [7]. There is evidence that stress decreases the ability to experience NACC mediated reward. Uncontrollable footshock specifically decreases the ICSS rate in the NACC immediately after and even 7 days after application of the stressor, however, without affecting ICSS rate in the SN [8]. Desmethylinipramine, a tricyclic antidepressant, reverses this footshock-induced decrease in ICSS rate in the NACC [146]. Similar findings were observed after immunological stress: injection with sheep red blood cells in mice, an antigen that induces a peak immune response at the fourth day after inoculation, reduced the response rate for ICSS in the NACC on the third, the fourth and the fifth day after inoculation. The dose of sheep red blood cells is known to influence DA activity in the NACC. Another stressor, food deprivation, had no effect on the response rate for ICSS in the NACC. This is quite remarkable since the NACC is known to be involved in food intake [54] (see also below: Food intake). In other brain areas, like the MFB [99], the LH and the substantia innominata [95] ICSS rate clearly increased after food deprivation. Finally, it was observed that the effect of stress on ICSS in the NACC depends on the mouse strain [148]. While there was a decrease in ICSS rate in DBA/2J mice after footshock stress, an increase in ICSS rate was observed in BALB/cByJ mice and no change in C57BL/6Jmice.

#### **Effect of drugs or brain lesions on ICSS in the NACC**

One way to study the mechanism of ICSS in the NACC is to administer pharmacological agents of different classes and to evaluate the effect on ICSS. For a summary of the findings of the literature we refer to Table 2.

#### *CNS stimulants*

Cocaine, amphetamine and methylenedioxymethamphetamine (MDMA) are three stimulants of the central nervous system (CNS), which all induce a significant increase in DA metabolism in the NACC (Table 1). After administration of these stimulants, rats will start ICSS in the NACC at lower stimulation frequencies and will press more and at higher rates [84, 90, 149]. Only for MDMA, a derivative of amphetamine commonly known as ecstasy, a decrease in the total number of presses was observed. MDMA affects the serotonergic system and

Table 1. *Drug effects on ICSS in the NACC*

Drug	Species	Admin. route	Dose	Effect	Ref.
<i>CNS stimulants</i>					
Cocaine	Wistar rat	IP	5 mg/kg 15 mg/kg	↑ ICSS rate & maximal rate, ↓ stimulation frequency threshold	[59]
– Mainly blocks the uptake of DA, 5-HT and NA at the neuronal plasma membrane transporters [116]; rewarding properties are mediated via simultaneous actions on DA, 5-HT and NA transporters [116]; 0.5 and 1 mg/kg systemic cocaine increases DA in the NACC shell [87]					
Amphetamine	Wistar rat	SC	100 µg	↑ ICSS rate	[118]
– release of monoamines (especially DA and NA) from nerve terminals; 0.5–3 mg/kg amphetamine increases DA in the NACC 7–25-fold [52, 88]; 1–3 mg/kg reduces 3,4-dihydroxyphenylacetic acid (DOPAC) and homovanillic acid (HVA) in the NACC with more than 50% [52]; no effect on 5-HT levels in the NACC, except at high dose (9 mg/kg)					
	Wistar rat	IP	0.3 mg/kg 1 mg/kg	↑ ICSS rate; highest dose: ↑ maximal rate & ↓ stimulation threshold	[59]
	Charles River rats	IP	1 mg/kg	↑ ICSS rate, also increase in function of time	[90]
+ 6-OH-DA lesion					
	Wistar rat	IP	1 mg/kg	↑ ICSS rate	[84]
	Wistar rat	LH	not applicable	lesions neutralized the effect of amphetamine	[84]
MDMA	Wistar rat	IP	0.5 mg/kg 2 mg/kg 4 mg/kg	highest dose: ↓ ICSS total lever presses & stimulation threshold	[59]
– binds to DA, 5-HT and NA transporters and reverses the action of these transporters, resulting in the release of DA, 5-HT and NA in the synapse; 1–3 mg/kg induces a 3 fold increase of DA and 5-HT in the NACC [52]; 1–3 mg/kg increases DOPAC in the NACC by 75–80% [52]; 3 mg/kg decreased the 5-HT metabolite 5-HIAA by 60% [52]					
+ Methysergide	Wistar rat	IP	5 mg/kg	reversal of MDMA induced decrease in ICSS rate & maximal rate	[59]
antagonist at 5-HT <sub>2</sub> receptor and agonist at some 5-HT <sub>1</sub> receptors [108]					
+ Ketanserin	Wistar rat	IP		no effect on MDMA induced decrease	[59]
5-HT <sub>2</sub> receptor antagonist					
Methysergide	Wistar rat	IP	5 mg/kg	no effect	[59]
<i>D2-like DA antagonists</i>					
Haloperidol	Wistar rat	SC	5 µg	↓ ICSS rate	[118]
– D2-like DA antagonist with α-adrenergic receptor affinity; 0.1–1 mg/kg increases DA release in the NACC by 50% [16, 58]					
	Rhesus monkey	IM	0.1–0.4 mg/kg (1*/2d) during 3 wks	ICSS at 25–75% lower stimulation amplitudes. This is reversible.	[104]
	Wistar rat	IP	0.07 mg/kg 0.2 mg/kg 0.67 mg/kg	↓ ICSS rate	[76]
+ Hyoscine	Wistar rat	IP	0.3 mg/kg 1.5 mg/kg	attenuation of haloperidol induced decrease in ICSS rate	
muscarinic acetylcholine receptor antagonist; 0.5 mg/kg increases DA in the NACC [45]					
Spiroperidol	Albino rat	NACC homo-lateral	1 µg	58% ↓ ICSS rate (range 25–83%)	[69]
– D2-like DA antagonist					
		NACC contra-lateral	1 µg	no effect	

(continued)

Table 1 (continued)

Drug	Species	Admin. route	Dose	Effect	Ref.
<i>Opioids and endorphins</i>					
Naltrexone – $\mu$ opioid receptor antagonist [122]; lower affinity for $\delta$ and $\kappa$ receptors, able to reverse agonists at $\delta$ and $\kappa$ sites [122]; 1 mg/kg systemic naltrexone had no effect on basal DA in the NACC but reverses ethanol or food intake induced DA release in the NACC [5, 112]	Long-Evans rat	IP	2.5 mg/kg 5 mg/kg 10 mg/kg 20 mg/kg	↓ ICSS rate & ↑ stimulation frequency necessary to obtain the same ICSS rate, not dose dependent	[123]
(Des-Tyr <sup>1</sup> )- $\gamma$ -endorphin – endogenous non-opioid peptide which probably acts on presynaptic mesolimbic dopamine receptors [120]; DT $\gamma$ E has no effect on basal DA release in the NACC in vitro but suppressed K <sup>+</sup> -induced DA release [102]	Wistar rat	SC	2.5 $\mu$ g/kg 25 $\mu$ g/kg	↓ ICSS rate, dose dependent	[118]
$\alpha$ -endorphin – endogenous opioid peptide; 10 and 20 $\mu$ g intracerebral $\alpha$ -endorphin tended to decrease DA and DOPAC in the striatum and 20 $\mu$ g decreased HVA [51]	Wistar rat	SC	2.5 $\mu$ g/kg 25 $\mu$ g/kg	no effect	[118]
<i>Tricyclic antidepressant</i>					
Desipramine – tricyclic antidepressant; inhibition of NA re-uptake; 5 mg/kg systemic desipramine has no effect on DA, DOPAC or HVA in the NACC [83]; in vitro has no effect on DA release in the NACC induced by electrical stimulation in the NACC [48]	CD-1 mice	IP	5 mg/kg	no effect on ICSS rate when given alone, reversal of the reduction in ICSS induced by uncontrollable footshocks	[145] [146]

Summary of drug effects on ICSS in the NACC, including drug dose and administration route. A decrease in ICSS threshold indicates that lower current intensities are needed to induce the same ICSS rate. *ICSS* Intracranial self-stimulation; *IM* intramuscular; *IP* intraperitoneal; *LH* lateral hypothalamus; *NACC* nucleus accumbens; *SC* subcutaneous

Table 2. Lesion effects on ICSS in the NACC

Lesion type	Species	Target	Effect	Reference
6-OH-DA lesion	Wistar rat	LH	↓ ICSS rate at 1–3 days after lesion, no effect during next 18 sessions	[84]
RF lesion	?	VTA	↑ ICSS rate	[109]
Blockade		GABAergic input to the mesolimbic DA neurons	↑ ICSS rates of the NACC	personal communication [139]

Summary of the effects of lesions in the brain on ICSS in the NACC, including the type and the target of the lesion. *6-OH-DA* 6-hydroxydopamine; *DA* dopamine; *GABA*: gamma-aminobutyric acid; *ICSS* intracranial self-stimulation; *LH* lateral hypothalamus; *NACC* nucleus accumbens; *RF* radiofrequency; *SC* subcutaneous; *VTA* ventral tegmental area

often induces motor deficits known as the '5-HT syndrome' [59]. The decrease in lever pressing is probably due to these motor deficits: concomitant administration of MDMA and methysergide, a 5-HT<sub>2</sub> antagonist and 5-HT<sub>1</sub> agonist which prevents these motor deficits, increased the total number of presses and ICSS rate [59]. Single methysergide administration and augmentation of MDMA with ketanserin, a drug related to methysergide but without 5-HT<sub>1</sub> agonistic properties, had no

effect. Therefore, it was concluded that the decrease in lever pressing with MDMA was due to 5-HT<sub>1</sub> receptor mediated motor deficits.

#### D2-like DA antagonists

Acute administration of haloperidol and spiroperidol, two D2-like DA antagonists, decreases the ICSS rate in the NACC in rats [67, 76, 118]. Although haloperidol

may induce disturbances in motor performance [44], it is unlikely that motor deficits induced the decrease in ICSS rate. Indeed, injection of spiroperidol in the NACC only decreased the ICSS rate when it was on the same side of the ICSS electrode. The lack of effect on ICSS when injected contralateral to the ICSS electrode suggests that motor performance was intact [69]. The decrease in ICSS rate induced by haloperidol could be attenuated by administration of hyoscine (scopolamine), which is a muscarinic acetylcholine receptor antagonist [76]. Contrasting these results, one study reported that ICSS was facilitated after chronic haloperidol administration: monkeys started ICSS at lower stimulation amplitudes [104].

#### *Opioids and endorphins*

The effect of naltrexone, (Des-Tyr<sup>1</sup>)- $\gamma$ -endorphin (DT $\gamma$ E) and  $\alpha$ -endorphin on ICSS in the NACC was verified. None of these agents influences basal DA release in the NACC but naltrexone and DT $\gamma$ E decrease evoked DA release in the NACC. It was observed that these two agents suppress ICSS in the NACC [118, 123]. In contrast,  $\alpha$ -endorphin, has no effect on ICSS in the NACC.

#### *Tricyclic antidepressant*

Desipramine (desmethylinipramine) had no effect on ICSS in the NACC but could reverse the reduction in ICSS induced by uncontrollable footshocks [145, 146].

#### *Brain lesions*

The NACC receives a massive DAergic projection from the VTA, which is part of the mesolimbic projection. Interruption of this pathway with a 6-hydroxydopamine (6-OH-DA) lesion at the level of the LH, leads to a decrease in ICSS rate after three days [84]. After long-term recovery (21 days), however, this decrease in ICSS rate normalised and even tended to increase [84]. Indeed, Simon *et al.* demonstrated an increase in ICSS in rats with a radiofrequency lesion in the VTA after 17 days [109]. The discrepancy between the short- and long-term effects of a lesion in the afferent DAergic projection, might be attributed to increased sensitivity for DA in the NACC due to upregulation of postsynaptic DA receptors [31]. Probably therefore, no correlation was observed between the increase in ICSS rate and the decrease in DA concentration [109].

#### *Dopamine*

The studies on the effects of drugs and brain lesions on ICSS in the NACC, present evidence for the involvement of the DAergic system. Agents, which increase DA release in the NACC, are likely to increase the ICSS rate or lower its threshold and vice versa. Additional evidence comes from fast cyclic voltammetry (FCV) studies. DA is one of the neurotransmitters that oxidises after application of a voltage waveform. The resulting current flow can be measured and is proportional to the DA concentration at the microrecording electrode. With this technique, called FCV, an increase in extracellular fluid DA at a distance of 200–400  $\mu$ m from the NACC stimulation electrode was measured *in vitro*. Local application of cocaine [125] facilitated DA release in the NACC induced by ES [10]. Other findings oppose the involvement of DA. Prado-Alcázar and Wise mapped ICSS sites in different regions in the brain but discovered no close correspondence between the boundaries of the reward system and those of the DA terminal fields as revealed by DA fluorescence [89, 147]. However, ICSS in the medial and ventral NACC, corresponding to regions of DA and cholecystokinin (CCK) co-localization, was accompanied with significant elevations in motor activity [147].

### **Other behavioural effects of ES in the NACC**

#### *Activity*

A significant increase in activity was reported following ES in the NACC in freely moving animals [32, 40] and in animals tested for ICSS [144], especially when stimulated in the medial NACC [147]. In addition, ES in the NACC influences changes in activity induced by the administration of different drugs. It enhanced the increase in activity after administration of amphetamine [56] and partly blocked the decrease in activity induced by a 5-HT<sub>1A</sub> agonist [117]. In contradiction to these increases in activity, stimulation in the NACC either had no effect [119] or even caused a decrease in activity in another study [93].

#### *Fight-or-flight*

Subjects facing threats dispose of a behavioural repertoire to handle the threat including opposing the threat (aggression or fighting) or flying away from it. The protagonists in the neurocircuitry involved in fight-or-flight behaviour are the AMY, the hypothalamus and the peri-



aqueductal gray [68]. The connectivity of the NACC with these brain areas suggests a role for the NACC as well in mediating these behavioural effects. Indeed, several experiments demonstrate that ES in the NACC can induce but also mitigate aggression and fighting behaviour, as well as fear.

Hano *et al.* observed violent running during NACC stimulation in rats, sometimes accompanied by backing, rearing on the hind paws, body shakes and increased muscular tone. Immediately after stimulation they noticed an increased excitement and aggression [40]. In accordance, ES in the NACC in male western fence lizards induced species-specific assertion display and challenge behaviour. In no case did stimulation elicit proper fighting in these lizards [114]. Two other reports, however, report the induction of aggression by ES in the LH and the decrease of this aggression by concomitant ES in the NACC [32, 33]. During ES in the LH, touching the region of the mouth elicits a biting reflex in cats. Stimulation at 6 or 60 Hz amplitude-dependently reduced the size of the region where these biting reflexes could be elicited. At the highest voltage level, the region for biting reflex was even completely abolished.

The effect of ES in the NACC on fear also varied considerably between studies. In one article, a decrease in fear responses was observed in cats during stimulation in the NACC [128], while in another an arrest reaction (sudden interruption of all movements) and escape behaviour were elicited at threshold and suprathreshold stimulation amplitudes, respectively [75]. In the latter study, the escape response consisted of movement of the cat to another place in the observation box, accompanied by crouching and flattening of the ears. It was suggested that the arrest-escape response was mediated via the efferent pathway to the VP, since a kainic lesion in this area increased the NACC stimulation threshold significantly (+51%). Finally, in fully conscious but restricted monkeys, no escape-like behaviour was observed [133].

#### *Exploratory behaviour*

An increase in sniffing was observed at 6 Hz stimulation in the NACC in cats. Above threshold voltage, stimulation caused sniffing, searching head movements, and in-and-out tongue movements [32]. In rats, continuous ES in the NACC also increased sniffing [40] and ICSS in the NACC increased normal sniffing [47] as well as amphetamine-induced sniffing [21]. ES during 10 days preceding the amphetamine injection had, how-

ever, no effect on amphetamine-induced sniffing [56]. Remarkably, in patients with OCD bilateral as well as unilateral ES produced a transient smell sensation. These olfactory perceptions were described as “something burning”, “stale air”, “an old bag”, “some sort of glue”, “something sweet”, or “as in nature”. One patient also tended to sniff the air in search of the source of the smell [27].

#### *Food intake*

Several interventions locally in the NACC influence feeding behaviour [151]. Upon ES in the NACC, increases as well as decreases in food intake were reported. In a food-reinforced task, food pellets were retrieved faster during ES in the NACC [119]. Also, upon termination of ICSS in the NACC, food intake was increased in normal rats, but decreased after amphetamine injections [21]. The decrease in the latter condition was larger than during amphetamine administration alone. Finally, a sudden interruption of all movements was observed in cats, including goal directed movements as those observed when the animals advance toward a dish of food [75].

ES resulted in considerable weight gain in 5 of 11 patients suffering from treatment refractory OCD (increase of 26, 13, 12, 12 and 8 kg). The increase in body weight is probably not just a consequence of the relief of OCD symptoms in these patients, since it was not always proportionate to the improvement in symptoms. Neither is it possible to state whether the increase in body weight is due to a stimulation-induced change in the subjects metabolism or hunger drive. Although patients were not asked to keep food diaries, some of them report to eat more and to crave for sweet things. Others deny eating more, but gain weight nevertheless. During consecutive episodes without stimulation, patients sometimes lost some of the gained weight. These effects of ES in the NACC on food intake are probably mediated by the connections of the NACC, mainly the mediodorsal shell, to the LH.

#### *Seizures*

Upon termination of stimulation, seizure like behaviour characterised by extreme hyperactivity, loud meowing, urination, and profuse salivation was observed in one cat (stimulation parameters: monophasic pulses, 0.5 ms pulse duration and 60 Hz frequency) [32]. In the ICSS experiments of Jenkins *et al.*, all rats stimulated in the NACC showed involuntary motor effects gradually

increasing during 3 weeks, comprising at first wet-dog shakes, eventually developing in full clonic seizures, which increased in frequency and severity (variable train duration, biphasic pulses, 0.2 ms pulse duration, 250–400  $\mu$ A, and 100 Hz frequency) [47]. In humans, no seizures were observed in the region of the anterior limbs of the internal capsule and the NACC, although stimulation was also performed with relative large pulse widths (0.06–0.45 ms) and at high frequency (100–130 Hz).

### Effects of ES in the NACC on compulsive behaviour

#### *Animal models of obsessive-compulsive disorder*

To examine whether ES in the NACC shell might benefit patients with treatment-refractory OCD, we electrically stimulated in the T-maze and the schedule-induced polydipsia animal model [117]. Spontaneous alternation behaviour is the natural tendency of most species to successively explore both arms of a T-maze alternately, provided the two goal boxes are equally reinforced. Subcutaneous injection of the selective 5-HT 1A receptor agonist 8-hydroxy-2-(di-n-propylamino)-tetralin hydrobromide reduces this alternation behaviour. This behaviour models the compulsive and repetitive behaviour of patients suffering from OCD [134]. In our experiments, we found that a lesion and ES in the NACC had the same behavioural effects in this model. However, alternation behaviour further decreased, suggesting that ES in the NACC would not benefit patients with OCD [117].

In the schedule-induced polydipsia model, hungry rats that receive a food pellet every minute will drink water after each pellet delivery, what also models compulsive and repetitive behaviour of human patients [132]. The total amount of water intake during the test sessions is inappropriate and even toxic especially when considering that the animals are in a deprived state (80% of normal body weight). In contrast to earlier findings in the T-maze model, we observed a stimulation amplitude-dependent reduction of schedule-induced polydipsia at high frequency but not at low frequency stimulation (unpublished personal observations). Finally, high-frequency stimulation in the NACC core decreased quinpirole-induced compulsive checking behaviour in rats [129]. In this animal model, quinpirole-sensitized rats return more often to their home cage in an open field, that models compulsive checking behaviour in patients suffering from obsessive-compulsive disorder [111].

#### *OCD patients*

In 1999, we implanted electrodes in the anterior limbs of the internal capsule of 4 treatment-resistant OCD patients with the most ventral contact (contact 0) being located near to or in the NACC. In three of them, marked beneficial effects could be demonstrated during acute test sessions [77]. In a subsequent blinded crossover design with randomly chronic stimulation on and off, a significant decrease on the Yale-Brown Obsessive Compulsive Scale and Clinical Global Severity scores was observed in 4 of 6 patients [78]. These stimulation-induced effects could be maintained for at least 21 months. Sturm *et al.* also demonstrated a favourable outcome of ES in the NACC in OCD and other anxiety disorders [100, 115]. Unipolar stimulation (90  $\mu$ s, 130 Hz, 2–6.5 V) in the right NACC resulted in a significant reduction of the symptoms in 3 of 4 patients. Bipolar stimulation was tried in one patient without additional improvement of the symptoms. In the fourth patient, a displacement of the electrode resulted in missing of the target area that might explain the negative outcome in this patient. In accordance, Lippitz *et al.* [60] showed that capsulotomies in the right hemisphere were decisive for a favourable therapeutic outcome.

### Physiological effects of ES in the NACC

Physiological parameters are likely to change in concert with behaviour. Nevertheless, we describe them in a separate section, since they were in some studies analysed under a general anaesthetic. Usually, ES in the NACC has only limited or inconsistent effects on physiological parameters.

#### *Cardiovascular effects*

ES in the NACC in freely-moving rats with parameters which induced ICSS, decreased the heart rate ( $-8.42 \pm 14.21$  beats/min; mean  $\pm$  SD) and increased the mean arterial pressure ( $9.5 \pm 7.7$  mmHg) [96]. Usually, the changes in mean arterial pressure preceded the changes in heart rate. During fear experiments, however, either no or inconsistent changes in heart rate were observed [128]. The cardiac responses upon ES in anaesthetized rats were also very limited. Ross and Malmö [96] found no changes in heart rate and a decrease of the mean arterial pressure in rats.

#### *Respiratory effects*

In a fear conditioning paradigm, ES in the NACC had little or no effect on respiration in awake cats except in

one of the three subjects in which an increase in respiration rate and a decrease in respiration amplitude were found [128]. In anaesthetized monkeys, there was no respiratory depression and no change in respiratory pattern or galvanic skin response [133]. In 6 of 8 patients with treatment-resistant OCD, switching the stimulator 'on' and 'off' induced a deep sigh. In addition, acute hyperventilation was observed in 4 of these patients when stimulated with particular contact combinations [27].

#### *Hormonal changes*

Koikegami *et al.* [55] found that ovulation can be induced in the unanaesthetized rabbit (a reflex-ovulating species) following a one-hour period of NACC stimulation. Also, oxytocin is released upon ES in the NACC [2]. In contrast, in female Wistar rats, ES in the NACC during 15 minutes had no significant effect on the concentration of plasma luteinising hormone although a slight elevation was present after 30 minutes [98]. Plasma cortisol and growth hormone levels were non-significantly increased in anaesthetized rhesus monkeys [25, 26, 133].

#### *Autonomic changes*

ES induced no autonomic effects like pupillary dilatation and salivation in one study [32] but increased alertness and induced pupillary dilatation and mild piloerection in another [75]. During ES in OCD patients, paresthesias or a warm feeling in certain body parts or over the whole body with transpiration and flushing were observed in all of them. Brusque abolition of stimulation frequently caused a transient hot feeling, transpiration and flushing as well [27].

### **Mechanisms of ES in the NACC**

ES is a rather new treatment option for patients with psychiatric disorders and is subject to improvement. Insight in the underlying mechanisms of ES may contribute to adaptations in the treatment. We already mentioned that FCV experiments indicate an increase in extracellular DA 200–400  $\mu\text{m}$  from the stimulation electrode in the NACC. Hence, stimulation in the NACC is likely to propagate DA release from DAergic afferents from the VTA and the SN. It is known that DA in the NACC is able to suppress spontaneous or glutamate-evoked firing of NACC neurons [124, 131, 140]. Other evidence for the mechanism of NACC ES comes from

electrophysiological experiments in which the effect of ES in the NACC on cellular activity in other brain targets was evaluated with microrecording. Neurons in brain targets lying downstream from the NACC may respond with excitation, inhibition, or a combination of both (early excitation followed by a period of inhibition and vice versa). Most neurons in the VP [20, 49, 64], the VTA [62, 138, 139] and the SN [101] were inhibited when single electrical pulses were given in the NACC, although excitation was also frequently observed in some studies [20, 57, 62]. The major part of the neurons in the AMY [66, 99] and the LH were excited with single pulse ES in the NACC [99]. Neurons in the tubero-infundibular hypothalamus were equally excited and inhibited [98] and the effect of stimulation in the NACC on neurons in the supraoptic nucleus depended on the cell type (excitation in vasopressin cells, inhibition in oxytocin cells) [107]. Finally, also neurons in the orbitofrontal cortex [94] and mediodorsal thalamic nucleus (MD) [70] responded orthodromically upon ES in the NACC. In addition to orthodromic activation, several authors reported antidromic neuronal activation in brain areas with afferents to the NACC like the prelimbic and orbitofrontal cortex [71, 94], agranular cortex [71], entorhinal cortex [24], HC [24, 136], AMY [66], MD [70], VTA [15, 29, 62, 69], SN [69, 101] and the VP [138]. It is clear from these studies that single pulse ES influences activity in several brain areas lying afferent and efferent to the NACC. Whereas low frequency stimulation may exert effects similar to single pulse stimulation, the effect of high frequency stimulation is likely to differ.

### **Discussion**

#### *High variability of behavioural effects*

The current overview of the behavioural changes demonstrates that ES in the NACC evokes a range of diverse effects. Depending on the publication even opposite observations were reported: ES in the NACC increased but also decreased aggressive behaviour, exploratory behaviour, food intake and compulsions in a model for OCD. In the next sections, we will discuss which factors might attribute to these contrasting results.

#### *Motivational context*

The effect of ES in the NACC may depend on the behavioural paradigm in which the animal was tested. Functionally, the NACC has been regarded as an inter-

face between motivation and action. Depending on the context and the motivational status of the subject, the response might differ. The NACC is anatomically well placed to accomplish this function. This nucleus receives its major afferents from the PFC (involved in higher functions like planning), the HC (memory and previous experiences) and the AMY (emotions), areas which belong to the limbic system, and projects towards the VP, the mesencephalon, both involved in motor behaviour, and the LH (e.g. food intake, ICSS, hormone regulation). Several of the afferents are able to gate information from other afferents by depolarising (or not) the membrane potential to an 'up' state (see Signal processing in the NACC).

#### *NACC subregional differences*

Histological staining and connectivity studies demonstrate that the NACC may be divided into a core and shell subregion (see: General features of the NACC). Connectivity patterns with other brain areas differ between and even within these subregions. Hence, depending on the electrode contact location, the behavioural and physiological effects may differ considerably. For instance, rostro-caudal, dorso-ventral and medio-lateral differences were observed between ICSS rates in the NACC [89, 147]. In these reports, high ICSS rates were observed in the ventral, caudal and medial NACC. It is likely that this subregion of the NACC with high rewarding properties is the most promising target for symptom relief with ES in humans suffering from psychiatric disorders in which depression is involved (like major depression disorder and obsessive-compulsive disorder with co-morbid depression). However, it is also possible that other structures, like the projections from the internal capsule or the nearby MFB mediate the good clinical effects of ES. The latter structure has higher rewarding properties (see above).

#### *Stimulation parameters*

The *amplitude* directly affects the extent of the stimulated region and the intensity of stimulation in nearby neurons.

The behavioural effects of *high versus low frequency* stimulation are often different and may be even opposite to each other. For instance, in Parkinson's disease, bradykinesia [73], tremor and the onset of myoclonic jerks [91] worsen at 5 Hz stimulation compared to stimulation with a frequency higher than 60 Hz, which leads to

symptom relief. In the NACC ICSS studies, response rates depended on the stimulation frequency (see above). There is neurophysiological evidence for the differential effects of high versus low frequency stimulation in the NACC. When recording single cell activities in the LH upon trains of electrical pulses at 50 Hz in the NACC, 16 neurons were excited and 8 neurons were inhibited (of the 31 LH neurons tested in total). Of the 8 inhibited neurons, 4 were also inhibited by single pulse stimulation whereas the other 4 and the 16 neurons, which were excited by stimulus trains, responded with excitation followed by inhibition [99]. The pulse width, on the other hand, directly influences the neuronal target element that is stimulated. Cell bodies and dendrites are optimally stimulated with a pulse width in the 1–10 ms range, small axons in the 200–700  $\mu$ s range and large myelinated axons in the 30–200  $\mu$ s range [43]. It is not clear whether behavioural differences in this review could be attributed to different pulse widths.

#### *Side effects*

Approximately, half of the OCD patients experienced weight gain during ES in the region of the NACC. This weight gain is probably not only due to symptom relief but also to a change in metabolism or an increase in hunger drive. From the ES experiments in animals, there is only limited evidence for physiological side effects. An increased blood pressure was observed in awake animals [96], which may increase the risk for cardiovascular diseases when it is sustained for a long period of time. However, the duration of this acute experiment was too short to take definite conclusions on increased blood pressure. Moreover, the raise in blood pressure may be related to behavioural changes induced by the ES. Seizures were observed in two animal studies in which large pulse widths and high stimulation frequency were used [32, 47]. Since stimulation in OCD patients is performed with similar stimulation parameters, the clinician has to be aware not to induce seizures when stimulating electrically in the NACC. The respiratory and hormonal changes were only of minor significance.

Finally, application of an electrical current in the brain may lead to an electrolytic lesion around the electrode tip, which depends on the electrical current and on electrode properties. First, the magnitude of the *charge density* around the electrode tip determines the susceptibility to induce a lesion. *Charge density* is the ratio of the energy per pulse over the free contact area of the electrode [65]. Second, biphasic stimulation is less det-



perimental than monophasic since every pulse is followed by a second pulse of the opposite polarity [97]. Third, platinum-iridium electrodes diminish the risk to induce an electrolytic lesion tenfold versus stainless steel electrodes [74]. For clinical applications, platinum-iridium electrodes are used. Post-mortem studies in chronically stimulated patients with intractable pain or movement disorders did not reveal electrolytic lesions around the electrode tip (e.g. Ref [3]). In most studies, there was limited gliosis around the foreign implanted material without any further implications. In one study, there was a limited lesion in a patient with Parkinson's disease probably due to migration of the electrode [42]. In this patient, the migration of the electrode was without side effects but the good clinical outcome of stimulation disappeared. In these post-mortem studies, none of the deaths was related to ES.

#### *ES in the NACC for OCD*

In the T-maze model and the schedule-induced polydipsia model, opposite findings were observed. In the former model, there was an increase in compulsive behaviour while there was a decrease in the latter. In humans, there is evidence from a limited number of patients that ES has a beneficial effect on the OCD symptoms. This puts the validity of the T-maze model into question. A good model for a psychiatric disorder has a highly predictive face, construct and discriminant validity [127]. The primary application of predictive validity is to assess the effects of potential therapeutic treatments: the model has *predictive validity* if it successfully discriminates between effective and ineffective treatments. *Face validity* concerns the degree to what extent a model resembles the condition being modelled. *Construct validity* means that the procedure in the model is based on a sound theoretical rationale. Finally, a model for OCD has *discriminant validity* when the evidence points to OCD as the disorder being modelled as distinct from a different or a non-specific psychiatric disorder [30]. In our experiments [117], the T-maze model incorrectly predicts that ES in the NACC would worsen the symptoms of OCD in humans. The face and construct validity of both models were not studied in our experiments but are discussed elsewhere [86].

#### *ES in the NACC for other psychiatric disorders*

In patients with treatment-resistant OCD, electrical stimulation in the region of the NACC significant-

ly decreases scores on the Hamilton Depression Scale (stimulation off: 26.7, stimulation on: 13.3) [27]. Therefore, a trial with electrical stimulation in the region of NACC was recently initiated in patients suffering from treatment-resistant major depression in our laboratory. Preliminary results are promising (own observations). Finally, electrical stimulation in the NACC may also be a potential new treatment in humans suffering from severe treatment-resistant addiction. The release of dopamine in the nucleus accumbens in humans is required in reward (e.g. drug high) and for the initiation of addiction [50]. Electrical stimulation in the NACC elicits DA release and is rewarding as well [125]. It is conceivable that certain stimulation parameters suppress or override addictive behavior. As far as we know the effect of ES in the NACC on addictive behavior has not been investigated in patients. However, a stereotactic lesion in the NACC reduced the relapse rate in patients suffering from addiction [28].

## Conclusion

ICSS experiments suggest that stimulation in the NACC has rewarding properties. Pharmacological studies point to the involvement of the DAergic system in mediating these rewarding effects. In addition, ES in the NACC affects general activity, fight-and-flight behaviour, exploration and food intake, although contrasting effects were often observed. In parallel with ES experiments in animal models for compulsions in OCD, a good clinical outcome was observed in patients with OCD during stimulation in the NACC.

## Acknowledgments

We acknowledge the financial support of the Research Fund K.U. Leuven (project VIS/02/007 and OT/03/57). This work has also been partially sponsored by a grant from Medtronic Europe.

## References

- Abbott A (2002) Brain implants show promise against obsessive disorder. *Nature* 419: 658
- Aulsebrook LH, Holland RC (1969) Central regulation of oxytocin release with and without vasopressin release. *Am J Physiol* 216: 818–829
- Baskin DS, Mehler WR, Hosobuchi Y, Richardson DE, Adams JE, Flitter MA (1986) Autopsy analysis of the safety, efficacy and cartography of electrical stimulation of the central gray in humans. *Brain Res* 371: 231–236
- Belleau ML, Warren RA (2000) Postnatal development of electrophysiological properties of nucleus accumbens neurons. *J Neurophysiol* 84: 2204–2216



5. Benjamin D, Grant ER, Pohorecky LA (1993) Naltrexone reverses ethanol-induced dopamine release in the nucleus accumbens in awake, freely moving rats. *Brain Res* 621: 137–140
6. Berendse HW, Groenewegen HJ (1990) Organization of the thalamostriatal projections in the rat, with special emphasis on the ventral striatum. *J Comp Neurol* 299: 187–228
7. Bland ST, Twining C, Watkins LR, Maier SF (2003) Stressor controllability modulates stress-induced serotonin but not dopamine efflux in the nucleus accumbens shell. *Synapse* 49: 206–208
8. Bowers WJ, Zacharko RM, Anisman H (1987) Evaluation of stressor effects on intracranial self-stimulation from the nucleus accumbens and the substantia nigra in a current intensity paradigm. *Behav Brain Res* 23: 85–93
9. Brog JS, Salyapongse A, Deutch AY, Zahm DS (1993) The patterns of afferent innervation of the core and shell in the “accumbens” part of the rat ventral striatum: immunohistochemical detection of retrogradely transported fluoro-gold. *J Comp Neurol* 338: 255–278
10. Brun P, Steinberg R, Le Fur G, Soubrie P (1995) Blockade of neurotensin receptor by SR 48692 potentiates the facilitatory effect of haloperidol on the evoked in vivo dopamine release in the rat nucleus accumbens. *J Neurochem* 64: 2073–2079
11. Buchanan SL, Thompson RH, Maxwell BL, Powell DA (1994) Efferent connections of the medial prefrontal cortex in the rabbit. *Exp Brain Res* 100: 469–483
12. Chang HT, Kitai ST (1985) Projection neurons of the nucleus accumbens: an intracellular labeling study. *Brain Res* 347: 112–116
13. Chiba T, Kayahara T, Nakano K (2001) Efferent projections of infralimbic and prelimbic areas of the medial prefrontal cortex in the Japanese monkey, *Macaca fuscata*. *Brain Res* 888: 83–101
14. Churchill L, Kalivas PW (1994) A topographically organized gamma-aminobutyric acid projection from the ventral pallidum to the nucleus accumbens in the rat. *J Comp Neurol* 345: 579–595
15. Clark D, Chiodo LA (1988) Electrophysiological and pharmacological characterization of identified nigrostriatal and mesoaccumbens dopamine neurons in the rat. *Synapse* 2: 474–485
16. De Deurwaerdere P, Moison D, Navailles S, Porras G, Spampinato U (2005) Regionally and functionally distinct serotonin receptors control in vivo dopamine outflow in the rat nucleus accumbens. *J Neurochem* 94: 140–149
17. DeFrance JF, Marchand JF, Sikes RW, Chronister RB, Hubbard JI (1985) Characterization of fimbria input to nucleus accumbens. *J Neurophysiol* 54: 1553–1567
18. DeFrance JF, Marchand JE, Stanley JC, Sikes RW, Chronister RB (1980) Convergence of excitatory amygdaloid and hippocampal input in the nucleus accumbens septi. *Brain Res* 185: 183–186
19. Deutch AY, Bourdelais AJ, Zahm DS (1993) The nucleus accumbens core and shell: accumbal compartments and their functional attributes. In: Kalivas PW, Barnes CD (eds) *Limbic motor circuits and neuropsychiatry*. CRC Press, pp 45–88
20. Dray A, Oakley NR (1978) Projections from nucleus accumbens to globus pallidus and substantia nigra in the rat. *Experientia* 34: 68–70
21. Eichler AJ, Antelman SM (1979) Sensitization to amphetamine and stress may involve nucleus accumbens and medial frontal cortex. *Brain Res* 176: 412–416
22. Fallon JH, Moore RY (1978) Catecholamine innervation of the basal forebrain. IV. Topography of the dopamine projection to the basal forebrain and neostriatum. *J Comp Neurol* 180: 545–580
23. Ferry AT, Ongur D, An X, Price JL (2000) Prefrontal cortical projections to the striatum in macaque monkeys: evidence for an organization related to prefrontal networks. *J Comp Neurol* 425: 447–470
24. Finch DM, Gigg J, Tan AM, Kosoyan OP (1995) Neurophysiology and neuropharmacology of projections from entorhinal cortex to striatum in the rat. *Brain Res* 670: 233–247
25. Frankel RJ, Jenkins JS, Wright JJ, Khan MU (1976) Effect of brain stimulation on aldosterone secretion in the rhesus monkey (*Macaca Mulatta*). *J Endocrinol* 71: 383–391
26. Frankel RJ, Jenkins JS, Wright JJ (1978) Pituitary-adrenal response to stimulation of the limbic system and lateral hypothalamus in the rhesus monkey (*Macaca mulatta*). *Acta Endocrinol (Copenh)* 88: 209–216
27. Gabriëls L (2004) Electrical brain stimulation in treatment refractory obsessive-compulsive disorder, University of Antwerpen
28. Gao G, Wang X, He S, Li W, Wang Q, Liang Q, Zhao Y, Hou F, Chen L, Li A (2003) Clinical study for alleviating opiate drug psychological dependence by a method of ablating the nucleus accumbens with stereotactic surgery. *Stereotact Funct Neurosurg* 81: 96–104
29. Gariano RF, Tepper JM, Sawyer SF, Young SJ, Groves PM (1989) Mesocortical dopaminergic neurons. I. Electrophysiological properties and evidence for soma-dendritic autoreceptors. *Brain Res Bull* 22: 511–516
30. Geyer MA, Markou A (1995) Animal models of psychiatric disorders. In: Bloom FE, Kupfer D (eds) *Psychopharmacology: fourth generation of progress*. Raven, New York, pp 787–798
31. Gnanalingham KK, Smith LA, Hunter AJ, Jenner P, Marsden CD (1993) Alterations in striatal and extrastriatal D-1 and D-2 dopamine receptors in the MPTP-treated common marmoset: an autoradiographic study. *Synapse* 14: 184–194
32. Goldstein JM, Siegel J (1980) Suppression of attack behavior in cats by stimulation of ventral tegmental area and nucleus accumbens. *Brain Res* 183: 181–192
33. Goldstein JM, Siegel J (1981) Stimulation of ventral tegmental area and nucleus accumbens reduce receptive fields for hypothalamic biting reflex in cats. *Exp Neurol* 72: 239–246
34. Gorelova N, Yang CR (1997) The course of neural projection from the prefrontal cortex to the nucleus accumbens in the rat. *Neuroscience* 76: 689–706
35. Goto Y, O'Donnell P (2001) Network synchrony in the nucleus accumbens in vivo. *J Neurosci* 21: 4498–4504
36. Greenberg BD, Price LH, Rauch SL, Friehs G, Noren G, Malone D, Carpenter LL, Rezai AR, Rasmussen SA (2003) Neurosurgery for intractable obsessive-compulsive disorder and depression: critical issues. *Neurosurg Clin N Am* 14: 199–212
37. Groenewegen HJ, Wright CI, Beijer AV, Voorn P (1999) Convergence and segregation of ventral striatal inputs and outputs. *Ann N Y Acad Sci* 877: 49–63
38. Haber SN, Lynd-Balta E, Mitchell SJ (1993) The organization of the descending ventral pallidal projections in the monkey. *J Comp Neurol* 329: 111–128
39. Haber SN, McFarland NR (1999) The concept of the ventral striatum in nonhuman primates. *Ann N Y Acad Sci* 877: 33–48
40. Hano J, Przewlocki R, Smialowska M, Chlapowska M, Rokosz-Pelc A (1978) The effect of electric stimulation of caudate nucleus and nucleus accumbens septi on serotonergic neurons in the rat brain. *Pol J Pharmacol Pharm* 30: 475–481
41. Heimer L, Zahm DS, Churchill L, Kalivas PW, Wohltmann C (1991) Specificity in the projection patterns of accumbal core and shell in the rat. *Neuroscience* 41: 89–125
42. Henderson JM, O'Sullivan DJ, Pell M, Fung VS, Hely MA, Morris JG, Halliday GM (2001) Lesion of thalamic centromedian – parafascicular complex after chronic deep brain stimulation. *Neurology* 56: 1576–1579
43. Holsheimer J, Demeulemeester H, Nuttin B, de Sutter P (2000) Identification of the target neuronal elements in electrical deep brain stimulation. *Eur J Neurosci* 12: 4573–4577
44. Hunt GE, McGregor IS (2002) Contrasting effects of dopamine antagonists and frequency reduction on Fos expression induced by lateral hypothalamic stimulation. *Behav Brain Res* 132: 187–201

45. Ichikawa J, Chung YC, Li Z, Dai J, Meltzer HY (2002) Cholinergic modulation of basal and amphetamine-induced dopamine release in rat medial prefrontal cortex and nucleus accumbens. *Brain Res* 958: 176–184
46. Jenike MA (1998) Neurosurgical treatment of obsessive-compulsive disorder. *Br J Psychiatry Suppl* 35: 79–90
47. Jenkins OF, Atrens DM, Jackson DM (1983) Self-stimulation of the nucleus accumbens and some comparisons with hypothalamic self-stimulation. *Pharmacol Biochem Behav* 18: 585–591
48. Jones SR, Garris PA, Kilts CD, Wightman RM (1995) Comparison of dopamine uptake in the basolateral amygdaloid nucleus, caudate-putamen, and nucleus accumbens of the rat. *J Neurochem* 64: 2581–2589
49. Jones DL, Mogenson GJ (1980) Nucleus accumbens to globus pallidus GABA projection: electrophysiological and iontophoretic investigations. *Brain Res* 188: 93–105
50. Kalivas PW, Volkow ND (2005) The neural basis of addiction: a pathology of motivation and choice. *Am J Psychiatry* 162: 1403–1413
51. Kameyama T, Ukai M, Noma S, Hiramatsu M (1982) Differential effects of alpha-, beta- and gamma-endorphins on dopamine metabolism in the mouse brain. *Brain Res* 244: 305–309
52. Kankaanpää A, Meririnne E, Lillsunde P, Seppälä T (1998) The acute effects of amphetamine derivatives on extracellular serotonin and dopamine levels in rat nucleus accumbens. *Pharmacol Biochem Behav* 59: 1003–1009
53. Kao CQ, Wang S (1985) Effect of stimulation of nucleus accumbens and naloxone microinjection on nociceptive unit discharges in the lateral habenular nucleus. *Sheng Li Xue Bao* 37: 24–30
54. Kelley AE (1999) Functional specificity of ventral striatal compartments in appetitive behaviors. *Ann N Y Acad Sci* 877: 71–90
55. Koikegami H, Hirata Y, Oguma J (1967) Studies on the paralimbic brain structures. *Folio Psych Neurol Jpn* 21: 151–180
56. Kokkinidis L, Kirkby RD, McCarter BD, Borowski TB (1989) Alterations in amphetamine-induced locomotor activity and stereotypy after electrical stimulation of the nucleus accumbens and neostriatum. *Life Sci* 44: 633–641
57. Lavin A, Grace AA (1996) Physiological properties of rat ventral pallidal neurons recorded intracellularly in vivo. *J Neurophysiol* 75: 1432–1443
58. Liegeois JF, Ichikawa J, Meltzer HY (2002) 5-HT<sub>2A</sub> receptor antagonism potentiates haloperidol-induced dopamine release in rat medial prefrontal cortex and inhibits that in the nucleus accumbens in a dose-dependent manner. *Brain Res* 947: 157–165
59. Lin HQ, Jackson DM, Atrens DM, Christie MJ, McGregor IS (1997) Serotonergic modulation of 3,4-methylenedioxymethamphetamine (MDMA)-elicited reduction of response rate but not rewarding threshold in accumbal self-stimulation. *Brain Res* 744: 351–357
60. Lippitz BE, Mindus P, Meyerson BA, Kihlstrom L, Lindquist C (1999) Lesion topography and outcome after thermocapsulotomy or gamma knife capsulotomy for obsessive-compulsive disorder: relevance of the right hemisphere. *Neurosurgery* 44: 452–458
61. Lopes da Silva FH, Arnolds DE, Neijt HC (1984) A functional link between the limbic cortex and ventral striatum: physiology of the subiculum accumbens pathway. *Exp Brain Res* 55: 205–214
62. Maeda H, Mogenson GJ (1980) An electrophysiological study of inputs to neurons of the ventral tegmental area from the nucleus accumbens and medial preoptic-anterior hypothalamic areas. *Brain Res* 197: 365–377
63. Mai J, Assheuer J, Paxinos G (2004) *Atlas of the human brain*, 2nd edn. Elsevier Academic Press, London
64. Maurice N, Deniau JM, Menetrey A, Glowinski J, Thierry AM (1997) Position of the ventral pallidum in the rat prefrontal cortex-basal ganglia circuit. *Neuroscience* 80: 523–534
65. McCreery DB, Agnew WF, Yuen TG, Bullara L (1990) Charge density and charge per phase as cofactors in neural injury induced by electrical stimulation. *IEEE Trans Biomed Eng* 37: 996–1001
66. Mello LE, Tan AM, Finch DM (1992) Convergence of projections from the rat hippocampal formation, medial geniculate and basal forebrain onto single amygdaloid neurons: an in vivo extra- and intracellular electrophysiological study. *Brain Res* 587: 24–40
67. Meyerson BA (1998) Neurosurgical treatment of mental disorders: introduction and indications. In: Gildenberg PL, Tasker RR (eds) *Textbook of stereotactic and functional neurosurgery*. McGraw Hill, New York, pp 1953–1963
68. Misslin R (2003) The defense system of fear: behavior and neurocircuitry. *Neurophysiol Clin* 33: 55–66
69. Mogenson GJ, Takigawa M, Robertson A, Wu M (1979) Self-stimulation of the nucleus accumbens and ventral tegmented area of Tsai attenuated by microinjections of spiroperidol into the nucleus accumbens. *Brain Res* 171: 247–259
70. Montaron MF, Buser P (1988) Relationships between nucleus medialis dorsalis, pericruciate cortex, ventral tegmental area and nucleus accumbens in cat: an electrophysiological study. *Exp Brain Res* 69: 559–566
71. Montaron MF, Deniau JM, Menetrey A, Glowinski J, Thierry AM (1996) Prefrontal cortex inputs of the nucleus accumbens-nigro-thalamic circuit. *Neuroscience* 71: 371–382
72. Morino P, Mascagni F, McDonald A, Hokfelt T (1994) Cholecystokinin corticostriatal pathway in the rat: evidence for bilateral origin from medial prefrontal cortical areas. *Neuroscience* 59: 939–952
73. Moro E, Esselink RJ, Xie J, Hommel M, Benabid AL, Pollak P (2002) The impact on Parkinson's disease of electrical parameter settings in STN stimulation. *Neurology* 59: 706–713
74. Mortimer JT, Shealy CN, Wheeler C (1970) Experimental non-destructive electrical stimulation of the brain and spinal cord. *J Neurosurg* 32: 553–559
75. Murer MG, Pazo JH (1993) Behavioral responses induced by electrical stimulation of the caudate nucleus in freely moving cats. *Behav Brain Res* 57: 9–19
76. Murzi E, Herberg LJ (1982) Anticholinergic treatment reverses haloperidol-induced blockade of self-stimulation of nucleus accumbens no less than of hypothalamus. *Q J Exp Psychol B* 34(Pt 1): 49–54
77. Nuttin B, Cosyns P, Demeulemeester H, Gybels J, Meyerson B (1999) Electrical stimulation in anterior limbs of internal capsules in patients with obsessive-compulsive disorder. *Lancet* 354: 1526
78. Nuttin BJ, Gabriels LA, Cosyns PR, Meyerson BA, Andriewitch S, Sunaert SG, Maes AF, Dupont PJ, Gybels JM, Gielen F, Demeulemeester HG (2003) Long-term electrical capsular stimulation in patients with obsessive-compulsive disorder. *Neurosurgery* 52: 1263–1272
79. O'Donnell P, Grace AA (1993) Physiological and morphological properties of accumbens core and shell neurons recorded in vitro. *Synapse* 13: 135–160
80. O'Donnell P, Grace AA (1995) Synaptic interactions among excitatory afferents to nucleus accumbens neurons: hippocampal gating of prefrontal cortical input. *J Neurosci* 15(5 Pt 1): 3622–3639
81. O'Donnell P, Greene J, Pabello N, Lewis BL, Grace AA (1999) Modulation of cell firing in the nucleus accumbens. *Ann N Y Acad Sci* 877: 157–175
82. Olds J, Milner P (1954) Positive reinforcement produced by electrical stimulation of septal area and other regions of rat brain. *J Comp Physiol Psychol* 47: 419–427
83. Pallis E, Thermos K, Spyrali C (2001) Chronic desipramine treatment selectively potentiates somatostatin-induced dopamine release in the nucleus accumbens. *Eur J Neurosci* 14: 763–767

84. Phillips AG, Fibiger HC (1978) The role of dopamine in maintaining intracranial self-stimulation in the ventral tegmentum, nucleus accumbens, and medial prefrontal cortex. *Can J Psychol* 32: 58–66
85. Phillipson OT, Griffiths AC (1985) The topographic order of inputs to nucleus accumbens in the rat. *Neuroscience* 16: 275–296
86. Pitman RK (1989) Animal models of compulsive behavior. *Biol Psychiatry* 26: 189–198
87. Pontieri FE, Tanda G, Di Chiara G (1995) Intravenous cocaine, morphine, and amphetamine preferentially increase extracellular dopamine in the “shell” as compared with the “core” of the rat nucleus accumbens. *Proc Natl Acad Sci USA* 92: 12304–12308
88. Porras G, Di Matteo V, Fracasso C, Lucas G, De Deurwaerdere P, Caccia S, Esposito E, Spampinato U (2002) 5-HT<sub>2A</sub> and 5-HT<sub>2C/2B</sub> receptor subtypes modulate dopamine release induced in vivo by amphetamine and morphine in both the rat nucleus accumbens and striatum. *Neuropsychopharmacology* 26: 311–324
89. Prado-Alcala R, Wise RA (1984) Brain stimulation reward and dopamine terminal fields. I. Caudate-putamen, nucleus accumbens and amygdala. *Brain Res* 297: 265–273
90. Predy PA, Kokkindis L (1984) Sensitization to the effects of repeated amphetamine administration on intracranial self-stimulation: evidence for changes in reward processes. *Behav Brain Res* 13: 251–259
91. Rizzone M, Lanotte M, Bergamasco B, Tavella A, Torre E, Facciani G, Melcarne A, Lopiano L (2001) Deep brain stimulation of the subthalamic nucleus in Parkinson's disease: effects of variation in stimulation parameters. *J Neurol Neurosurg Psychiatry* 71: 215–219
92. Robinson TG, Beart PM (1988) Excitant amino acid projections from rat amygdala and thalamus to nucleus accumbens. *Brain Res Bull* 20: 467–471
93. Rolls ET (1971) Contrasting effects of hypothalamic and nucleus accumbens septi self-stimulation on brain stem single unit activity and cortical arousal. *Brain Res* 31: 275–285
94. Rolls ET (1972) Activation of amygdaloid neurones in reward, eating and drinking elicited by electrical stimulation of the brain. *Brain Res* 45: 365–381
95. Rolls ET, Burton MJ, Mora F (1980) Neurophysiological analysis of brain-stimulation reward in the monkey. *Brain Res* 194: 339–357
96. Ross AR, Malmö RB (1979) Cardiovascular responses to rewarding brain stimulation. *Physiol Behav* 22: 1005–1013
97. Rowland V, MacIntyre WJ, Bidder TG (1960) The production of brain lesions with electrical current. II. Bidirectional currents. *J Neurosurg* 17: 55–69
98. Saphier DJ (1985) Nucleus accumbens and preoptic area stimulation: tuberoinfundibular single unit responses, modulation of electrical activity and gonadotrophin secretion. *Exp Brain Res* 57: 400–403
99. Sasaki K, Ono T, Muramoto K, Nishino H, Fukuda M (1984) The effects of feeding and rewarding brain stimulation on lateral hypothalamic unit activity in freely moving rats. *Brain Res* 322: 201–211
100. Saxena S, Rauch SL (2000) Functional neuroimaging and the neuroanatomy of obsessive-compulsive disorder. *Psychiatr Clin North Am* 23: 563–586
101. Scarnati E, Campana E, Pacitti C (1983) The functional role of the nucleus accumbens in the control of the substantia nigra: electrophysiological investigations in intact and striatum-globus pallidus lesioned rats. *Brain Res* 265: 249–257
102. Schoemaker H, Nickolson VJ (1980) Effects of des-Tyr-gamma-endorphin on dopamine release from various rat brain regions in vitro. *Life Sci* 27: 1371–1376
103. Schuurman PR, Bosch DA, Bossuyt PM, Bonsel GJ, van Someren EJ, de Bie RM, Merkus MP, Speelman JD (2000) A comparison of continuous thalamic stimulation and thalamotomy for suppression of severe tremor. *N Engl J Med* 342: 461–468
104. Seeger TF, Gardner EL (1979) Enhancement of self-stimulation behavior in rats and monkeys after chronic neuroleptic treatment: evidence for mesolimbic supersensitivity. *Brain Res* 175: 49–57
105. Sesack SR, Deutch AY, Roth RH, Bunney BS (1989) Topographical organization of the efferent projections of the medial prefrontal cortex in the rat: an anterograde tract-tracing study with Phaseolus vulgaris leucoagglutinin. *J Comp Neurol* 290: 213–242
106. Sesack SR, Pickel VM (1990) In the rat medial nucleus accumbens, hippocampal and catecholaminergic terminals converge on spiny neurons and are in apposition to each other. *Brain Res* 527: 266–279
107. Shibuki K (1984) Supraoptic neurosecretory cells: synaptic inputs from the nucleus accumbens in the rat. *Exp Brain Res* 53: 341–348
108. Silberstein SD (1998) Methysergide. *Cephalalgia* 18: 421–435
109. Simon H, Stinus L, Tassin JP, Tassin JP, Lavielle S, Blanc G, Thierry AM, Glowinski J, Le Moal M (1979) Is the dopaminergic mesocorticolimbic system necessary for intracranial self-stimulation? Biochemical and behavioral studies from A10 cell bodies and terminals. *Behav Neural Biol* 27: 125–145
110. Sturm V, Lenartz D, Koulousakis A, Treuer H, Herholz K, Klein JC, Klosterkötter J (2003) The nucleus accumbens: a target for deep brain stimulation in obsessive-compulsive- and anxiety-disorders. *J Chem Neuroanat* 26: 293–299
111. Szechtman H, Eckert MJ, Tse WS, Boersma JT, Bonura CA, McClelland JZ, Culver KE, Eilam D (2001) Compulsive checking behavior of quinpirole-sensitized rats as an animal model of Obsessive-Compulsive Disorder (OCD): form and control. *BMC Neurosci* 2: 4
112. Taber MT, Zernig G, Fibiger HC (1998) Opioid receptor modulation of feeding-evoked dopamine release in the rat nucleus accumbens. *Brain Res* 785: 24–30
113. Takagishi M, Chiba T (1991) Efferent projections of the infralimbic (area 25) region of the medial prefrontal cortex in the rat: an anterograde tracer PHA-L study. *Brain Res* 566: 26–39
114. Tarr RS (1982) Species typical display behavior following stimulation of the reptilian striatum. *Physiol Behav* 29: 615–620
115. Tass PA, Klosterkötter J, Schneider F, Lenartz D, Koulousakis A, Sturm V (2003) Obsessive-compulsive disorder: development of demand-controlled deep brain stimulation with methods from stochastic phase resetting. *Neuropsychopharmacology* 28 Suppl 1: S27–S34
116. Uhl GR, Hall FS, Sora I (2002) Cocaine, reward, movement and monoamine transporters. *Mol Psychiatry* 7: 21–26
117. van Kuyck K, Demeulemeester H, Feys H, De Weerd W, Dewil M, Tousseyn T, De Sutter P, Gybels J, Bogaerts K, Dom R, Nuttin B (2003) Effects of electrical stimulation or lesion in nucleus accumbens on the behaviour of rats in a T-maze after administration of 8-OH-DPAT or vehicle. *Behav Brain Res* 140: 165–173
118. Van Ree JM, Otte AP (1980) Effects of (Des-Tyr<sup>1</sup>)-gamma-endorphin and alpha-endorphin as compared to haloperidol and amphetamine on nucleus accumbens self-stimulation. *Neuropharmacology* 19: 429–434
119. Velley L, Cardo B (1979) Long-term improvement of learning after early electrical stimulation of some central nervous structures: is the effect structure and age-dependent? *Brain Res Bull* 4: 459–466
120. Verhoef JC, Scholtens H, Vergeer EG, Witter A (1985) Des-Tyr<sup>1</sup>-gamma-endorphin (DT gamma E) and des-enkephalin-gamma-endorphin (DE gamma E): plasma profile and brain uptake after systemic administration in the rat. *Peptides* 6: 467–474

121. Walaas I, Fonnum F (1979) The distribution and origin of glutamate decarboxylase and choline acetyltransferase in ventral pallidum and other basal forebrain regions. *Brain Res* 177: 325–336
122. Way WL, Fields HL, Schumacher MA (2001) Opioid analgesics and antagonists. In: Katzung BG (ed) *Basic & Clinical Pharmacology*. Lange Medical Books/McGraw-Hill, pp 512–529
123. West TE, Wise RA (1988) Effects of naltrexone on nucleus accumbens, lateral hypothalamic and ventral tegmental self-stimulation rate-frequency functions. *Brain Res* 462: 126–133
124. White FJ, Wang RY (1986) Electrophysiological evidence for the existence of both D-1 and D-2 dopamine receptors in the rat nucleus accumbens. *J Neurosci* 6: 274–280
125. Wiczorek W, Kruk ZL (1995) Influences of neuronal uptake and D2 autoreceptors on regulation of extracellular dopamine in the core, shell and rostral pole of the rat nucleus accumbens. *Brain Res* 699: 171–182
126. Willner P (1984) The validity of animal models of depression. *Psychopharmacology (Berl)* 83: 1–16
127. Willner P (1986) Validation criteria for animal models of human mental disorders: learned helplessness as a paradigm case. *Prog Neuropsychopharmacol Biol Psychiatry* 10: 677–690
128. Wilson WJ (1983) Nucleus accumbens inhibits specific motor but not nonspecific classically conditioned responses. *Brain Res Bull* 10: 505–515
129. Winter C, Jalali R, Hosmann K, Kupsch A, Morgenstern R, Juckel G (2004) High frequency stimulation of the accumbens, the subthalamic nucleus, and the amygdala differentially affects quinpirole induced compulsive checking behavior in rats. *Society for Neuroscience, Washington, DC*, p 118
130. Wood PB (2004) Stress and dopamine: implications for the pathophysiology of chronic widespread pain. *Med Hypotheses* 62: 420–424
131. Woodruff GN, McCarthy PS, Walker RJ (1976) Studies on the pharmacology of neurones in the nucleus accumbens of the rat. *Brain Res* 115: 233–242
132. Woods A, Smith C, Szewczak M, Dunn RW, Cornfeldt M, Corbett R (1993) Selective serotonin re-uptake inhibitors decrease schedule-induced polydipsia in rats: a potential model for obsessive compulsive disorder. *Psychopharmacology (Berl)* 112: 195–198
133. Wright J, Kelly D, Mitchell-Heggs N, Frankel R (1977) Respiratory changes induced by intracranial stimulation: anatomical localizing value and related functional effects in rhesus monkeys. In: Sweet WH, Obrador S, Martin-Rodriguez JG (eds) *Neurosurgical treatment in psychiatry, pain, and epilepsy*. University Park Press, Baltimore, pp 751–756
134. Yadin E, Friedman E, Bridger WH (1991) Spontaneous alternation behavior: an animal model for obsessive-compulsive disorder? *Pharmacol Biochem Behav* 40: 311–315
135. Yang CR, Mogenson GJ (1984) Electrophysiological responses of neurones in the nucleus accumbens to hippocampal stimulation and the attenuation of the excitatory responses by the mesolimbic dopaminergic system. *Brain Res* 324: 69–84
136. Yang CR, Mogenson GJ (1986) Dopamine enhances terminal excitability of hippocampal-accumbens neurons via D2 receptor: role of dopamine in presynaptic inhibition. *J Neurosci* 6: 2470–2478
137. Yang CR, Mogenson GJ (1989) Ventral pallidal neuronal responses to dopamine receptor stimulation in the nucleus accumbens. *Brain Res* 489: 237–246
138. Yim CY, Mogenson GJ (1980) Electrophysiological studies of neurons in the ventral tegmental area of Tsai. *Brain Res* 181: 301–313
139. Yim CY, Mogenson GJ (1980) Effect of picrotoxin and nipectic acid on inhibitory response of dopaminergic neurons in the ventral tegmental area to stimulation of the nucleus accumbens. *Brain Res* 199: 466–473
140. Yim CY, Mogenson GJ (1982) Response of nucleus accumbens neurons to amygdala stimulation and its modification by dopamine. *Brain Res* 239: 401–415
141. Yim CY, Mogenson GJ (1988) Neuromodulatory action of dopamine in the nucleus accumbens: an in vivo intracellular study. *Neuroscience* 26: 403–415
142. Zaborszky L, Alheid GF, Beinfeld MC, Eiden LE, Heimer L, Palkovits M (1985) Cholecystokinin innervation of the ventral striatum: a morphological and radioimmunological study. *Neuroscience* 14: 427–453
143. Zaborszky L, Cullinan WE (1992) Projections from the nucleus accumbens to cholinergic neurons of the ventral pallidum: a correlated light and electron microscopic double-immunolabeling study in rat. *Brain Res* 570: 92–101
144. Zacharko RM, Bowers WJ, Kokkinidis L, Anisman H (1983) Region-specific reductions of intracranial self-stimulation after uncontrollable stress: possible effects on reward processes. *Behav Brain Res* 9: 129–141
145. Zacharko RM, Bowers WJ, Anisman H (1984) Responding for brain stimulation: stress and desmethylinpramine. *Prog Neuropsychopharmacol Biol Psychiatry* 8: 601–606
146. Zacharko RM, Bowers WJ, Kelley MS, Anisman H (1984) Prevention of stressor-induced disturbances of self-stimulation by desmethylinpramine. *Brain Res* 321: 175–179
147. Zacharko RM, Kasian M, Irwin J, Zalzman S, LaLonde G, MacNeil G, Anisman H (1990) Behavioral characterization of intracranial self-stimulation from mesolimbic, mesocortical, nigrostriatal, hypothalamic and extra-hypothalamic sites in the non-inbred CD-1 mouse strain. *Behav Brain Res* 36: 251–281
148. Zacharko RM, Lalonde GT, Kasian M, Anisman H (1987) Strain-specific effects of inescapable shock on intracranial self-stimulation from the nucleus accumbens. *Brain Res* 426: 164–168
149. Zacharko RM, Zalzman S, Macneil G, Andrews M, Mendella PD, Anisman H (1997) Differential effects of immunologic challenge on self-stimulation from the nucleus accumbens and the substantia nigra. *Pharmacol Biochem Behav* 58: 881–886
150. Zaczek R, Hedreen JC, Coyle JT (1979) Evidence for a hippocampal-septal glutamatergic pathway in the rat. *Exp Neurol* 65: 145–156
151. Zahm DS (2000) An integrative neuroanatomical perspective on some subcortical substrates of adaptive responding with emphasis on the nucleus accumbens. *Neurosci Biobehav Rev* 24: 85–105
152. Zahm DS, Heimer L (1993) Specificity in the efferent projections of the nucleus accumbens in the rat: comparison of the rostral pole projection patterns with those of the core and shell. *J Comp Neurol* 327: 220–232

Correspondence: Bart Nuttin, Laboratory of Experimental Neurosurgery and Neuroanatomy, Department of Neuroscience and Psychiatry, K.U. Leuven Provisorium I, Minderbroedersstraat 17, 3000 Leuven, Belgium. e-mail: bart.nuttin@uz.kuleuven.ac.be